



## The gut microbiome in frail and non-frail elders

*Comprehensive Management of Aging in HIV infected subjects Workshop*

Nov, 25<sup>th</sup>, 2016

Marc Noguera-Julian, PhD

# Outline

- Human microbiome
- Aging & Frailty Syndrome
- Gut Microbiome in frail aging
- Gut Microbiome in extreme longevity
- Gut Microbiome in non-frail aging
- The FRAME Study
- Summary



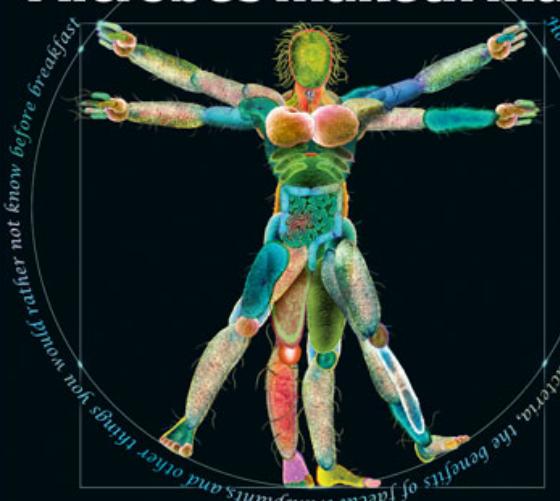
The Economist

AUGUST 18TH-24TH 2012

Economist.com

The Catholic church's unholy mess  
Paul Ryan: the man with the plan  
Generation Xhausted  
China, victim of the Olympics?  
On the origin of species

## Microbes maketh man



# nature

THE INTERNATIONAL WEEKLY JOURNAL OF SCIENCE

First results  
from the Human  
Microbiome  
Project highlight  
the healthy  
variation in  
our microbial  
selves

PAGES 194, 207 & 215



## FELLOW TRAVELLERS

ALS

GATHE  
SURE  
convincingly  
hydrogen

CLIMATE CHANGE

GET USED TO  
UNCERTAINTY  
Climate modelling  
faces its limits

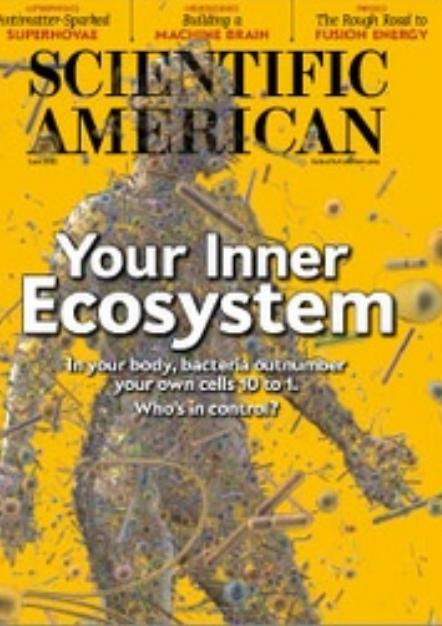
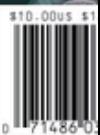
PAGE 183

SOLAR SYSTEM

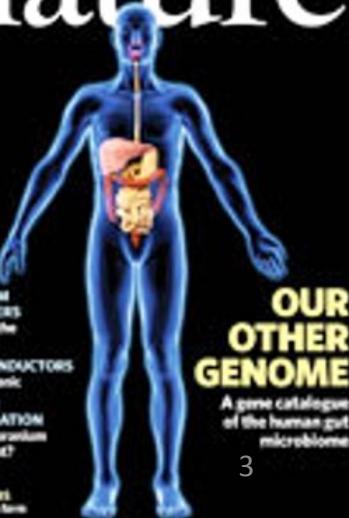
TITAN'S ELUSIVE  
METHANE  
Tropical lakes on Saturn's  
enigmatic moon?

PAGE 237

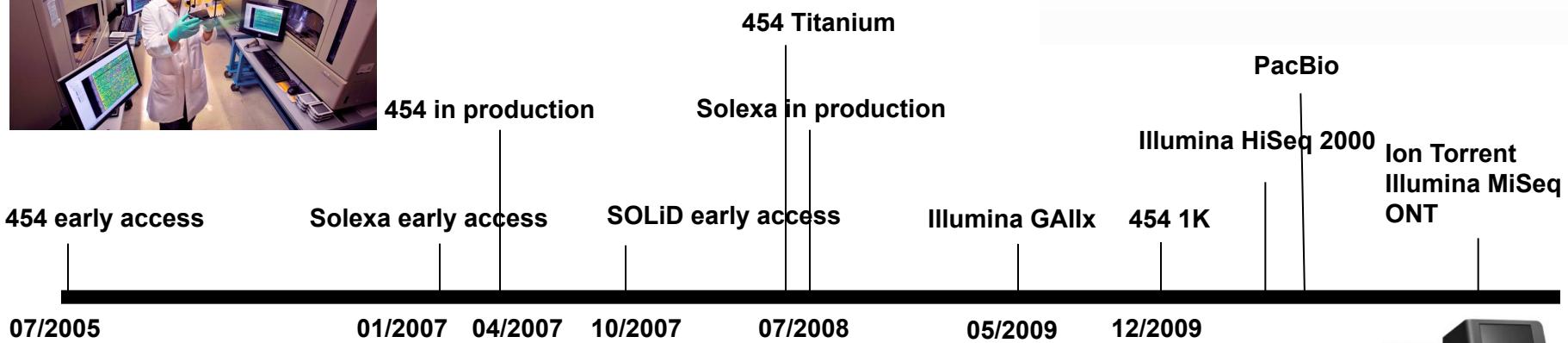
NATURE.COM  
14 June 2012



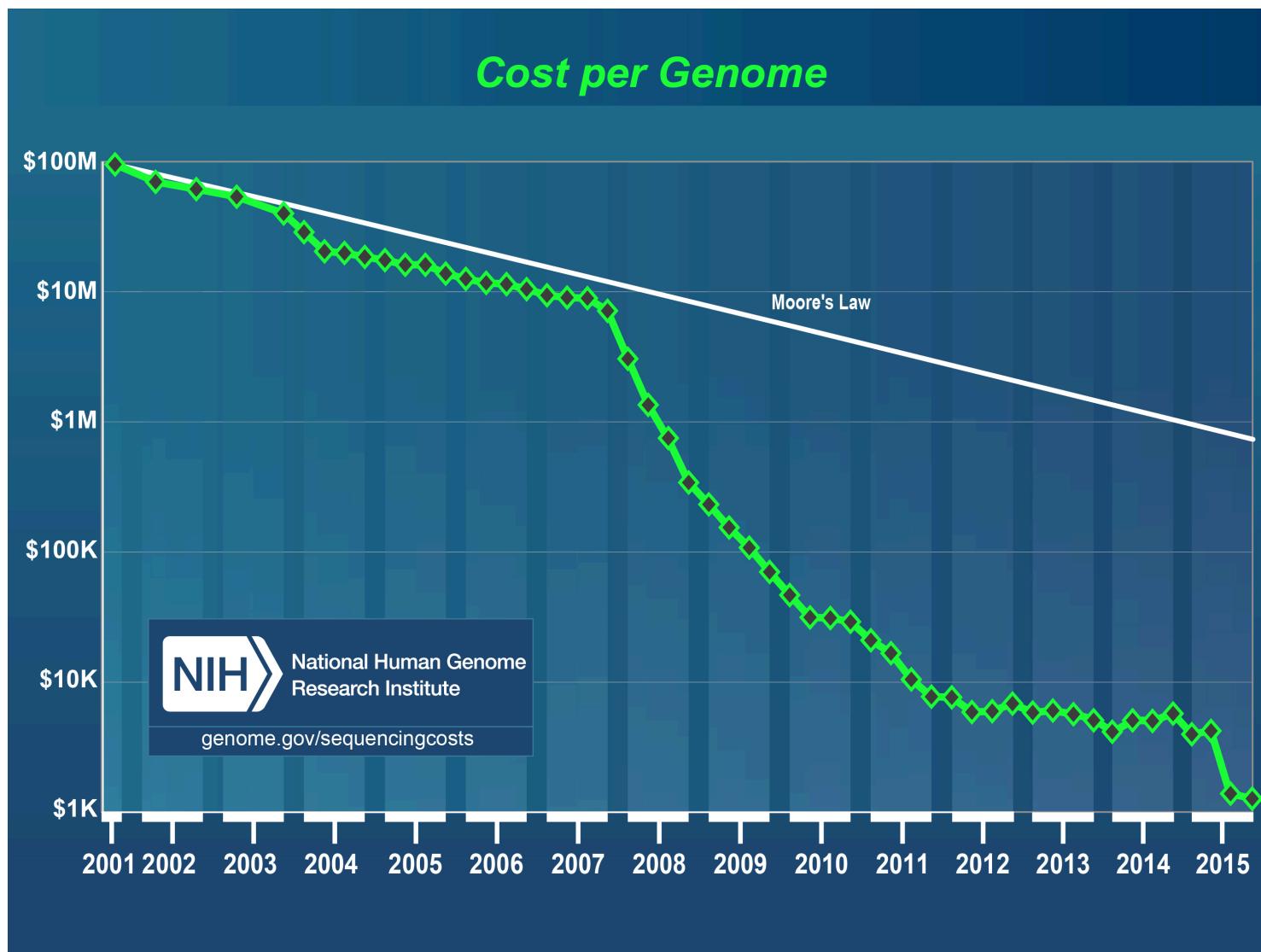
# nature

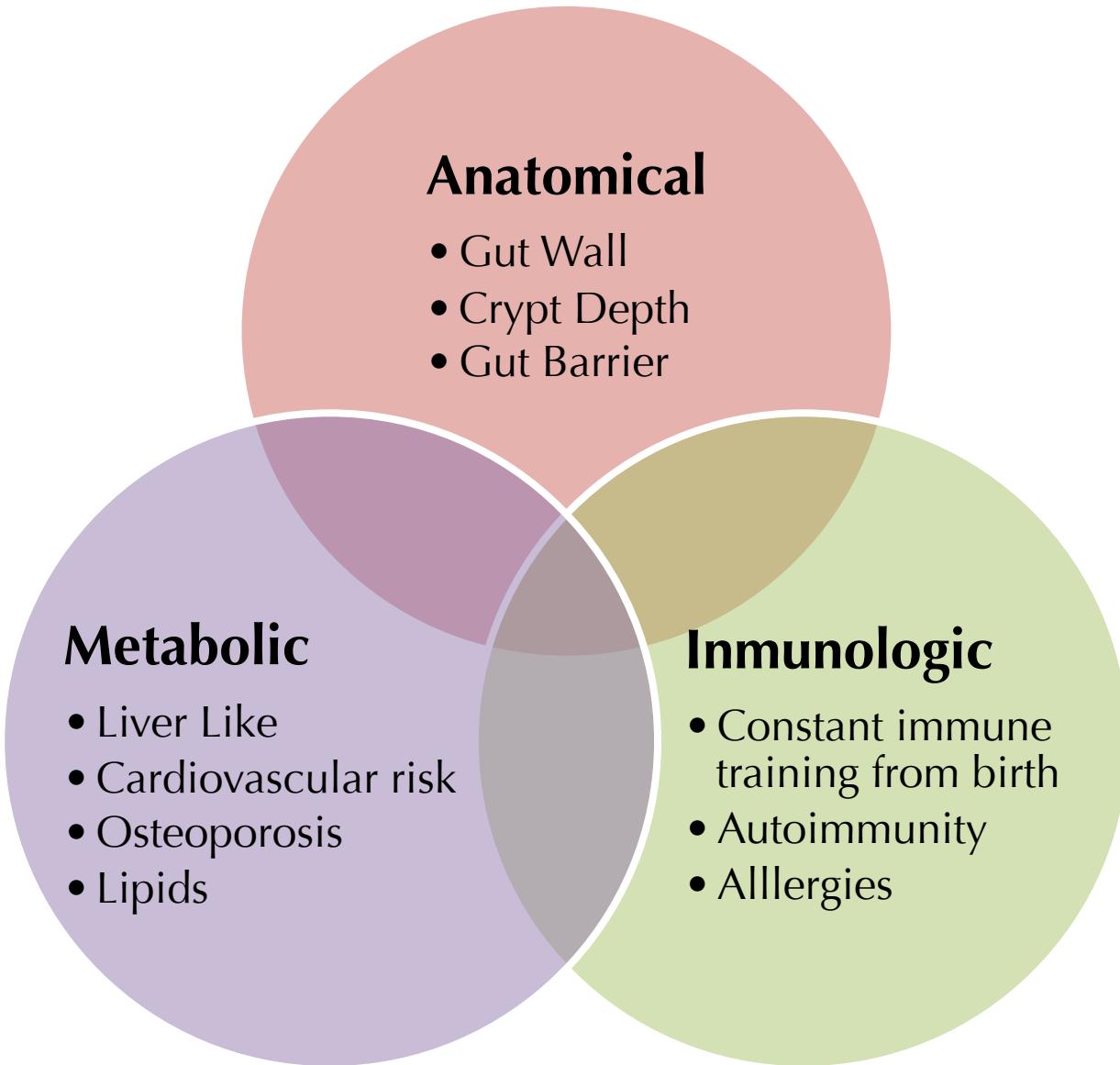


# The human microbiome – Next Generation Sequencing



# NGS Sequencing Costs

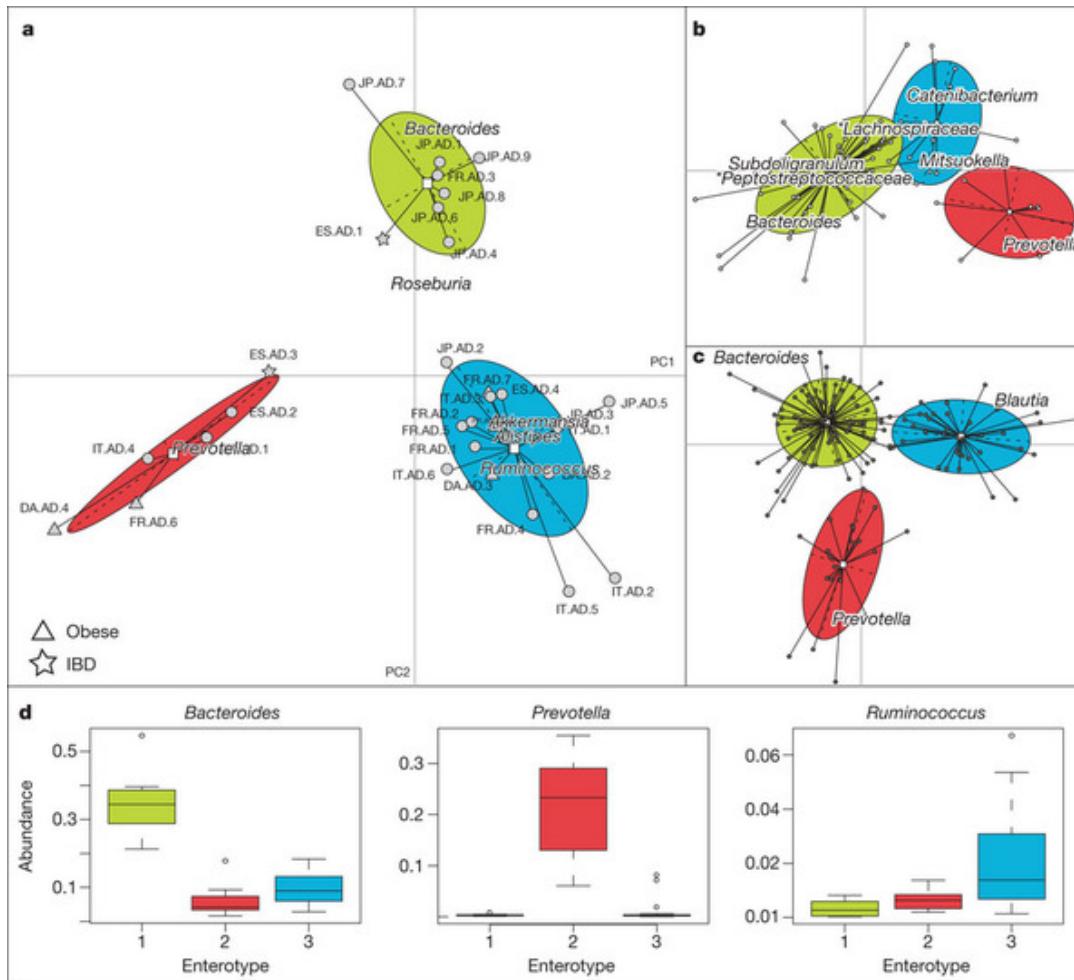


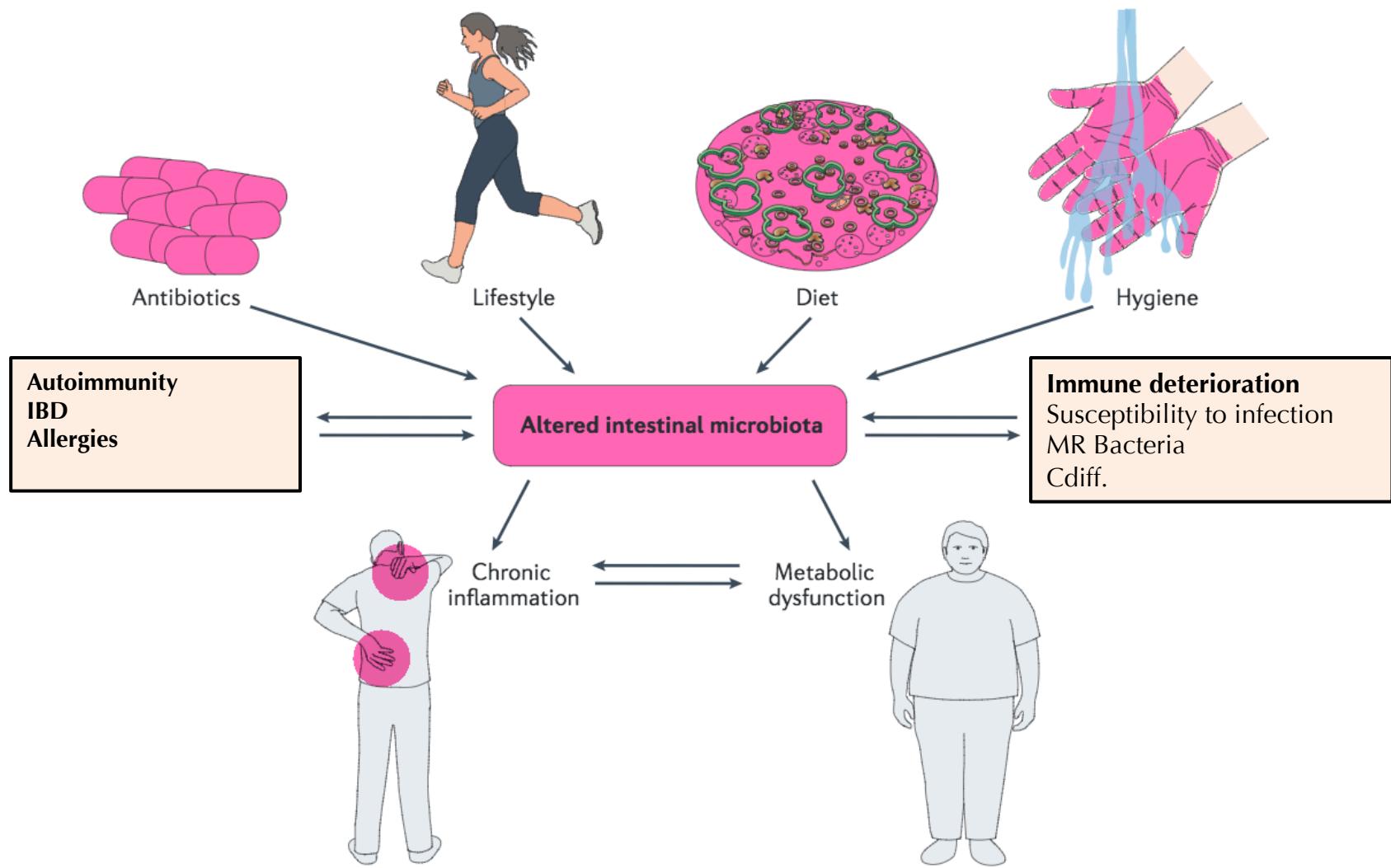


# Gut Microbiome's composition is structured

## Enterotypes of the human gut microbiome

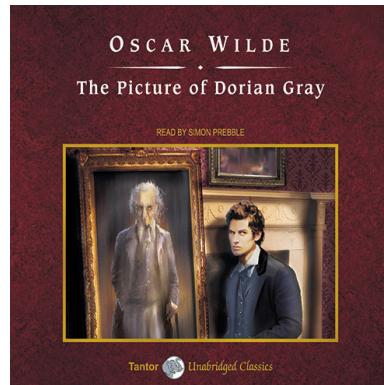
Manimozhiyan Arumugam<sup>1\*</sup>, Jeroen Raes<sup>1,2\*</sup>, Eric Pelletier<sup>3,4,5</sup>, Denis Le Paslier<sup>3,4,5</sup>, Takuji Yamada<sup>1</sup>, Daniel R. Mende<sup>1</sup>, Gabriel R. Fernandes<sup>1,6</sup>, Julien Tap<sup>1,7</sup>, Thomas Bruls<sup>3,4,5</sup>, Jean-Michel Batto<sup>7</sup>, Marcelo Bertalan<sup>8</sup>, Natalia Borruel<sup>9</sup>, Francesc Casellas<sup>9</sup>, Leyden Fernandez<sup>10</sup>, Laurent Gautier<sup>8</sup>, Torben Hansen<sup>11,12</sup>, Masahira Hattori<sup>13</sup>, Tetsuya Hayashi<sup>14</sup>, Michiel Kleerebezem<sup>15</sup>, Ken Kurokawa<sup>16</sup>, Marion Leclerc<sup>7</sup>, Florence Levenez<sup>7</sup>, Chaysavanh Manichanh<sup>9</sup>, H. Bjorn Nielsen<sup>8</sup>, Trine Nielsen<sup>11</sup>, Nicolas Pons<sup>7</sup>, Julie Poulaïn<sup>3</sup>, Junjie Qin<sup>17</sup>, Thomas Sicheritz-Ponten<sup>8,18</sup>, Sebastian Tims<sup>15</sup>, David Torrents<sup>10,19</sup>, Edgardo Ugarte<sup>3</sup>, Erwin G. Zoetendal<sup>15</sup>, Jun Wang<sup>17,20</sup>, Francisco Guarner<sup>9</sup>, Oluf Pedersen<sup>11,21,22,23</sup>, Willem M. de Vos<sup>15,24</sup>, Søren Brunak<sup>8</sup>, Joel Dore<sup>7</sup>, MetaHIT Consortium†, Jean Weissenbach<sup>3,4,5</sup>, S. Dusko Ehrlich<sup>7</sup> & Peer Bork<sup>1,25</sup>



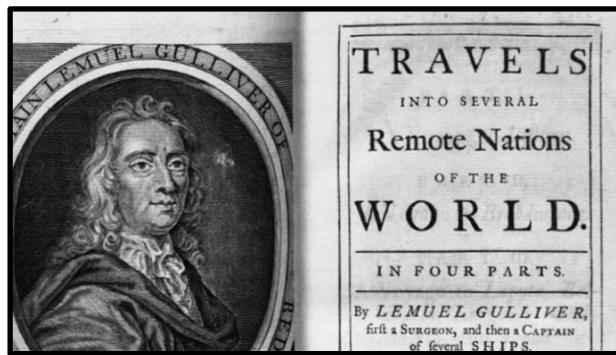


“In the land of frailty, confusion, contradiction, and ambiguity  
reign supreme.”

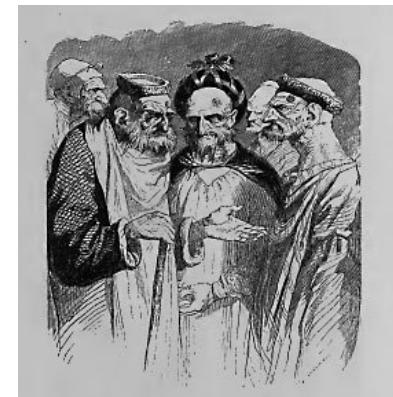
# Aging & Frailty



FRAILTY: “liability to be crushed or to decay, either in material or immaterial sense ... moral weakness, instability of mind, liability to err or yield to temptation”

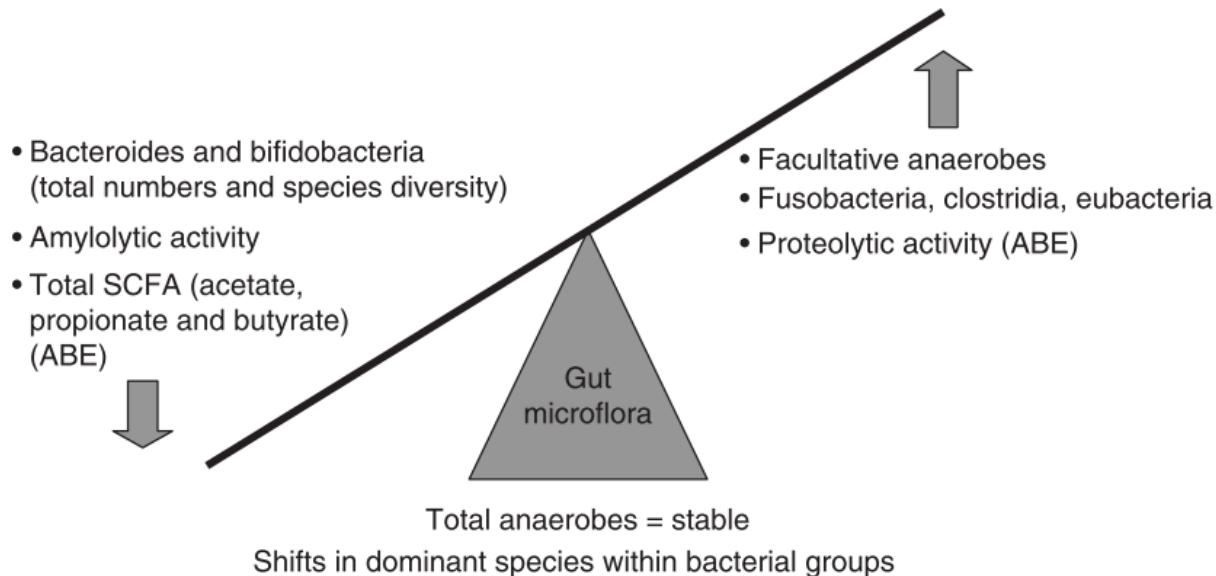


“when they come to fourscore years, which is reckoned the extremity of living in this country, they had not only all the follies and infirmities of other older men, but many more which arose from the dreadful prospect of never dying.”



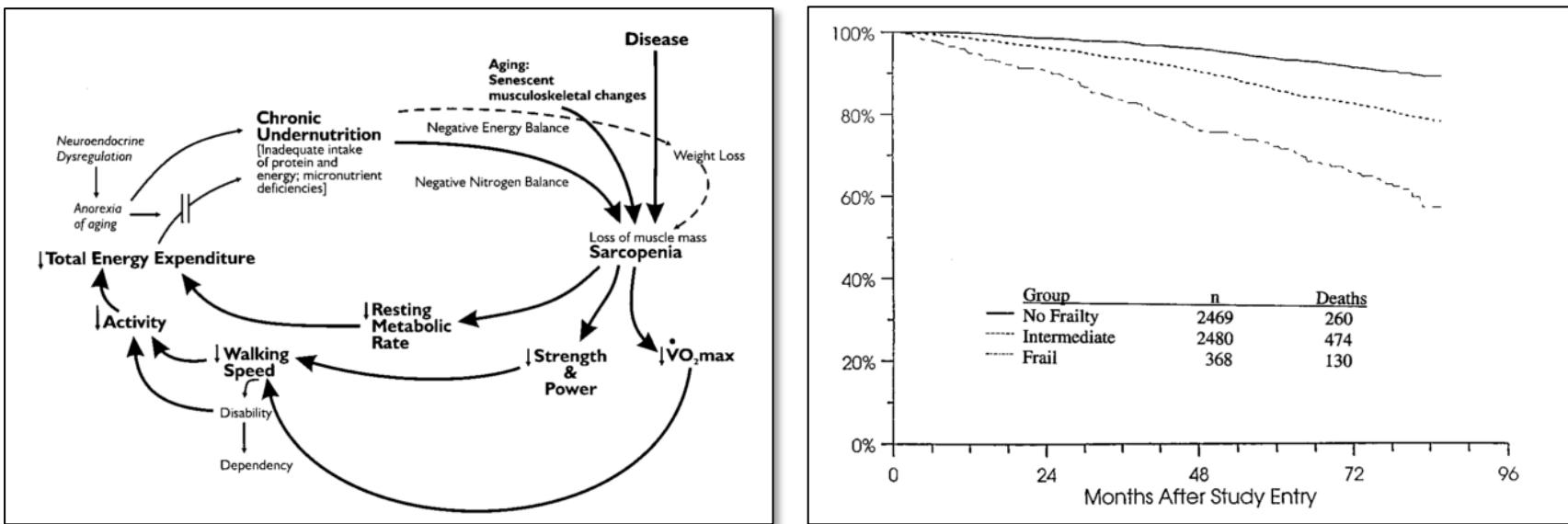
## Aging & Gut microbiome

- Changes in the GI tract(mastication function, loss of teeth, swallowing difficulties) can lead to nutritional imbalance
- Decreased intestinal motility and slow intestinal transit results in constipation and reduction of faecal bacterial matter with an associated increase of putrefactive processes.
- Age associated inflammation ('inflammaging') is a hallmark of aging and is linked to immune senescence and increased activation of several pro-inflammatory pathways.



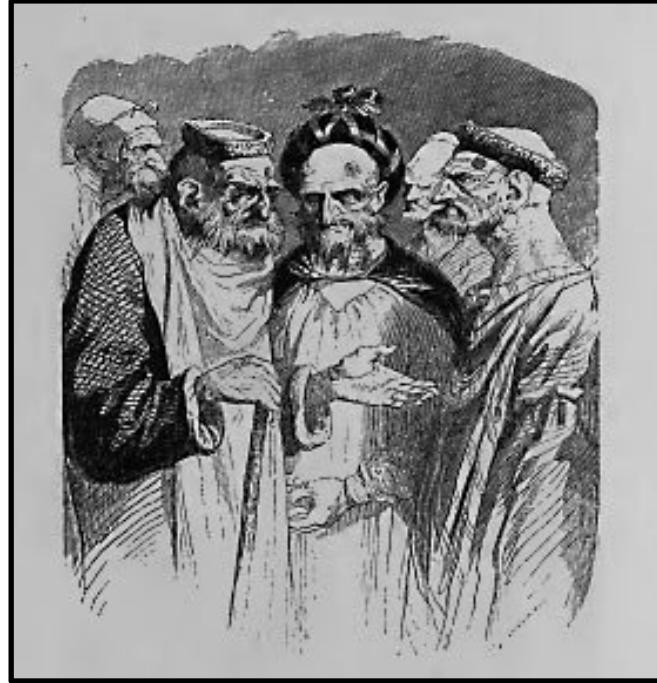
## FRAILTY

- Frailty is a complex multifunctional syndrome characterized by loss of weight & muscular mass, weakness, functionality loss within and aging background
- Functionally defined by deficit accumulation. Different Scales (Fried<sup>1,2</sup>, Edmonton<sup>3</sup>, SOF<sup>4</sup>...).
- Prevalence of frailty/Pre-frailty increases with age range from ~7-10% or 20-30% in >65 or >85 year old population.<sup>5</sup>
- Biologically characterized to a decreased function of innate and adaptive immune system, dysregulation of several key physiological systems, including the neuroendocrine, musculoskeletal, metabolic and immune/inflammatory<sup>6</sup> including increased levels of several inflammation markers(IL-6,CRP).



<sup>1</sup>Fried LP et al. Frailty in older adults: evidence for a phenotype. *J Gerontol A Biol Sci Med Sci* 2001, 56:M146-56. <sup>2</sup>Bandeen-Roche K, et al: Phenotype of frailty: characterization in the women's health and aging studies. *J Gerontol A Biol Sci Med Sci* 2006, 61:262-6. <sup>3</sup>Rolfsen DB, Majumdar SR, Tsuyuki RT, Tahir A, Rockwood K: Validity and reliability of the Edmonton Frail Scale. *Age Ageing* 2006, 35:526-9. <sup>4</sup>SOF index: 1. Ensor KE: Comparison of 2 Frailty Indexes for Prediction of Falls, Disability, Fractures, and Death in Older Women. *Arch Intern Med* 2008, 168:382. <sup>5</sup>Collard RM, Boter H, Schoevers RA, Oude Voshaar RC: Prevalence of frailty in community-dwelling older persons: a systematic review. *J Am Geriatr Soc* 2012, 60:1487-92. <sup>6</sup>Fulop T, McElhaney J, Pawelec G, Cohen AA, Morais JA, Dupuis G, Baehl S, Camous X, Witkowski JM, Larbi A: Frailty, Inflammation and Immunosenescence. *Interdisciplinary topics in gerontology and geriatrics* 2015:26-40.

The gut microbiome in frailty



**The Struldbrugs**  
Gulliver's Travels, by Jonathan Swift

# Microbiome in Frailty – Bartosch, 2004

Bartosch S, Fite A, Macfarlane GT, McMurdo MET: Characterization of bacterial communities in feces from healthy elderly volunteers and hospitalized elderly patients using real-time PCR and effects of antibiotic treatment on the fecal microbiota. *Appl Environ Microbiol* 2004, 70:9575–81.

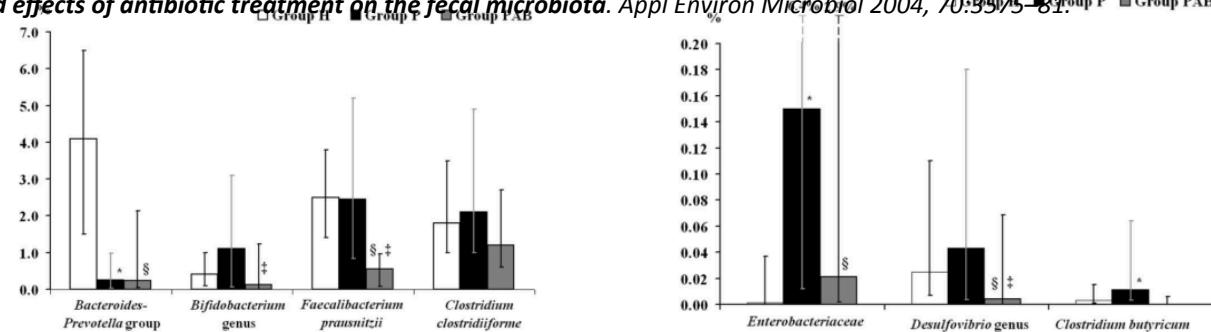


FIG. 1. Percentage of 16S rRNA gene copies of the predominant fecal bacterial groups and species in relation to total bacterial 16S rRNA gene copies (relative abundance). The percentage was calculated for each individual, and the median was determined for each subject group. A nonparametric Mann-Whitney U test was used for pairwise comparison of the subject groups. Error bars represent the interquartile range. \*, significant difference ( $P < 0.05$ ) between groups H and P; §, significant difference ( $P < 0.05$ ) between groups H and PAB; ‡, significant difference ( $P < 0.05$ ) between groups P and PAB.

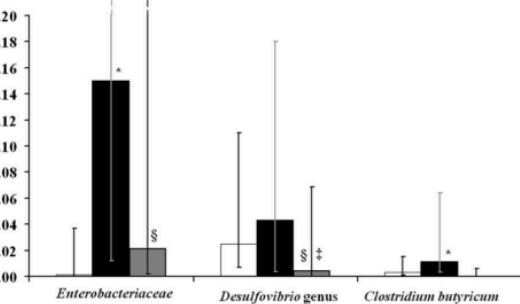


FIG. 2. Percentage of 16S rRNA gene copies of less abundant fecal bacterial groups and species in relation to total bacterial 16S rRNA gene copies (relative abundance). The percentage was calculated for each individual, and the median was determined for each subject group. A nonparametric Mann-Whitney U test was used for pairwise comparison of the subject groups. Error bars represent the interquartile range. \*, significant difference ( $P < 0.05$ ) between groups H and P; §, significant difference ( $P < 0.05$ ) between groups H and PAB; ‡, significant difference ( $P < 0.05$ ) between groups P and PAB.

TABLE 4. Summary of findings on fecal bacteria in groups of hospitalized patients compared to the control group

| Organism                            | Shifts observed from control group H to group P |                         |            | Shifts observed from control group H to group PAB |                          |            |
|-------------------------------------|---|-------------------------|------------|---|--------------------------|------------|
|                                     | 16S rRNA gene copy numbers                      | Relative abundance      | Prevalence | 16S rRNA gene copy numbers                        | Relative abundance       | Prevalence |
| All cubacteria                      | Significant reduction                           | Not applicable          | No change  | Significant reduction                             | Not applicable           | No change  |
| <i>Bacteroides-Prevotella</i> group | Significant reduction                           | Significant reduction   | Decrease   | Significant reduction                             | Significant reduction    | Decrease   |
| <i>Bifidobacterium</i> genus        | Nonsignificant reduction                        | Nonsignificant increase | No change  | Significant reduction                             | No change                | Decrease   |
| <i>Desulfovibrio</i> genus          | Nonsignificant reduction                        | Nonsignificant increase | No change  | Significant reduction                             | Significant reduction    | Decrease   |
| <i>Enterobacteriaceae</i>           | Nonsignificant increase                         | Significant increase    | No change  | No change   | Significant reduction    | No change  |
| <i>Faecalibacterium prausnitzii</i> | Significant reduction                           | No change               | No change  | Significant reduction                             | Significant reduction    | No change  |
| <i>Clostridium clostridiiforme</i>  | Significant reduction                           | No change               | No change  | Significant reduction                             | Nonsignificant reduction | No change  |
| <i>Clostridium butyricum</i>        | No change                                       | Significant increase    | No change  | Significant reduction                             | Significant reduction    | Decrease   |
| <i>Ruminococcus albus</i>           | Nonsignificant reduction                        | No change               | Decrease   | Significant reduction                             | Significant reduction    | Decrease   |
| <i>Enterococcus faecalis</i>        | Nonsignificant increase                         | Significant increase    | Increase   | Significant increase                              | Significant increase     | Increase   |

## Microbiome in Frailty – van Tongeren, 2005

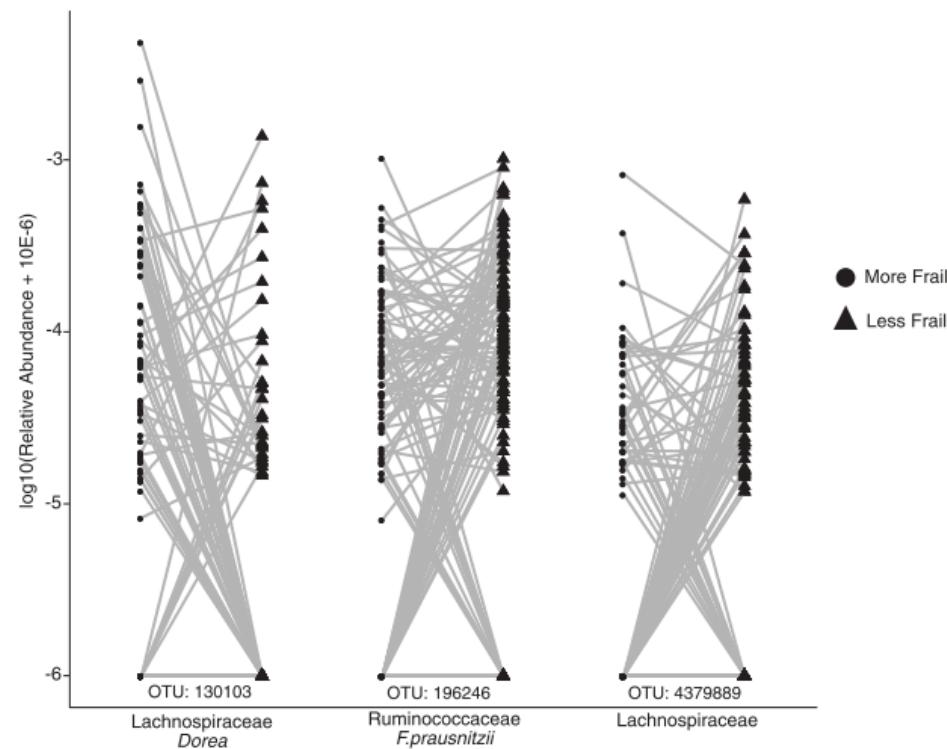
van Tongeren SP, Slaets JPJ, Harmsen HJM, Welling GW: *Fecal microbiota composition and frailty*. Appl Environ Microbiol 2005, 71:6438–42.

- Cross-sectional study from 23 elderly (median 86 years) in residential care
- Fluorescent in situ hybridization (FISH)
- GFI (Groningen) frailty scale determines
  
- Fecal samples from volunteers with high frailty scores showed a significant reduction in the number **of lactobacilli** (26-fold).
- At much higher population levels, both **the Bacteroides/Prevotella** (threefold) and the *Faecalibacterium prausnitzii* (fourfold) groups showed a significant reduction in percentage of total number of hybridizable bacteria in the elderly with high frailty scores.
- In contrast to this, the number of Enterobacteriaceae was significantly higher (sevenfold) in samples from very frail volunteers.

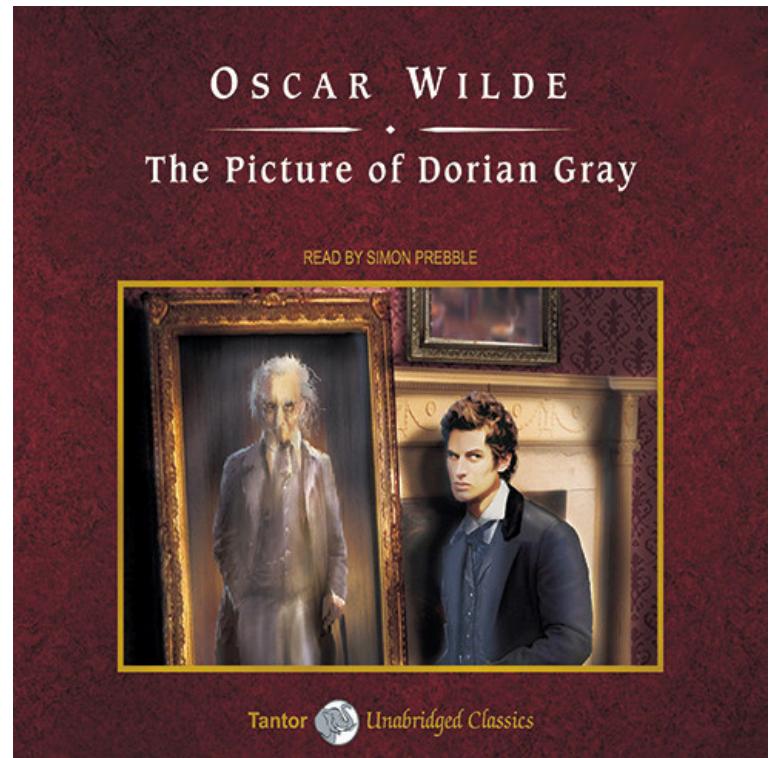
## Microbiome signatures in frailty – Jackson, 2016

Jackson M, et al: Signatures of early frailty in the gut microbiota. *Genome Med* 2016, 8:8.

- Twin Study including 728 female twins with (mean age 63 yo) from twinsUK cohort.
- 16s rRNA sequencing from fecal samples
- Frailty quantified through the Rockwood Frailty Index. Generally low frailty indices.
- Frailty negatively associated with gut microbiota diversity
- *F. Prausnitzii* and *Ruminococcaceae* negatively correlated and *E. dolichum* and *E. lenta*.
- *E. lenta* has been associated to GI disease while *E. dolichum* is enriched in genes involved in simple sugar processing



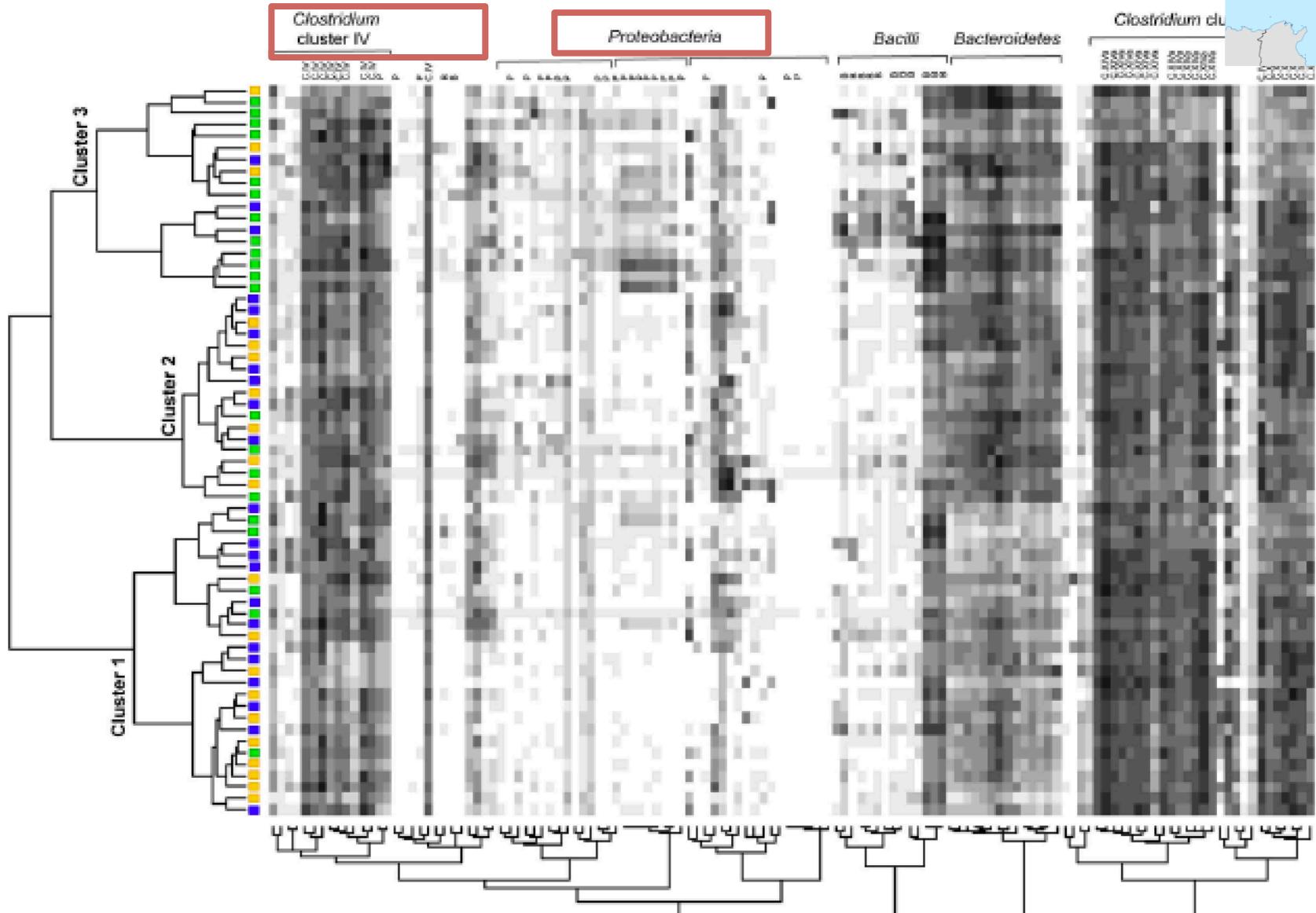
## The microbiome in extreme longevity



## Microbiome in Centenarian subjects – Biagi, 2010

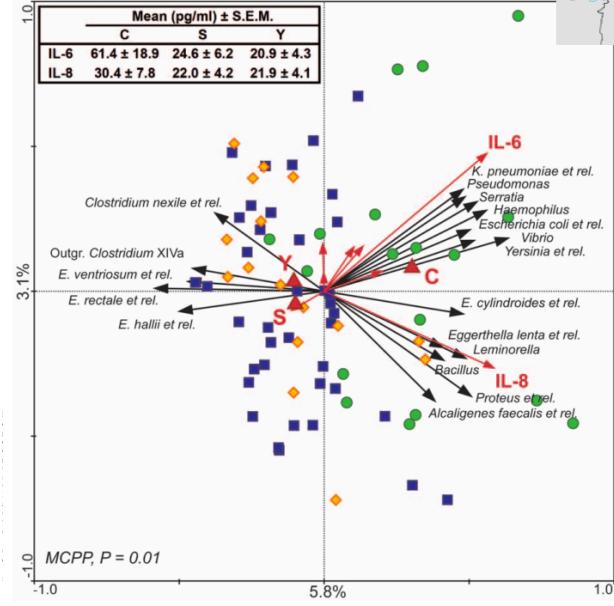
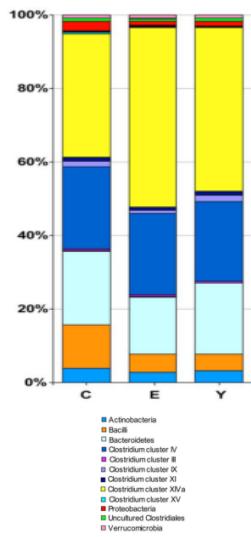
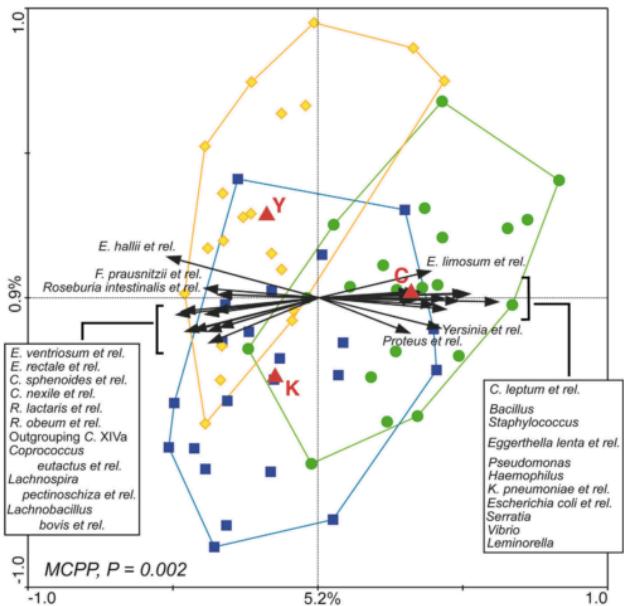


Biagi E, et al: *Through ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians*. PLoS One 2010, 5:e10667.



# Microbiome in Centenarian subjects – Biagi, 2010

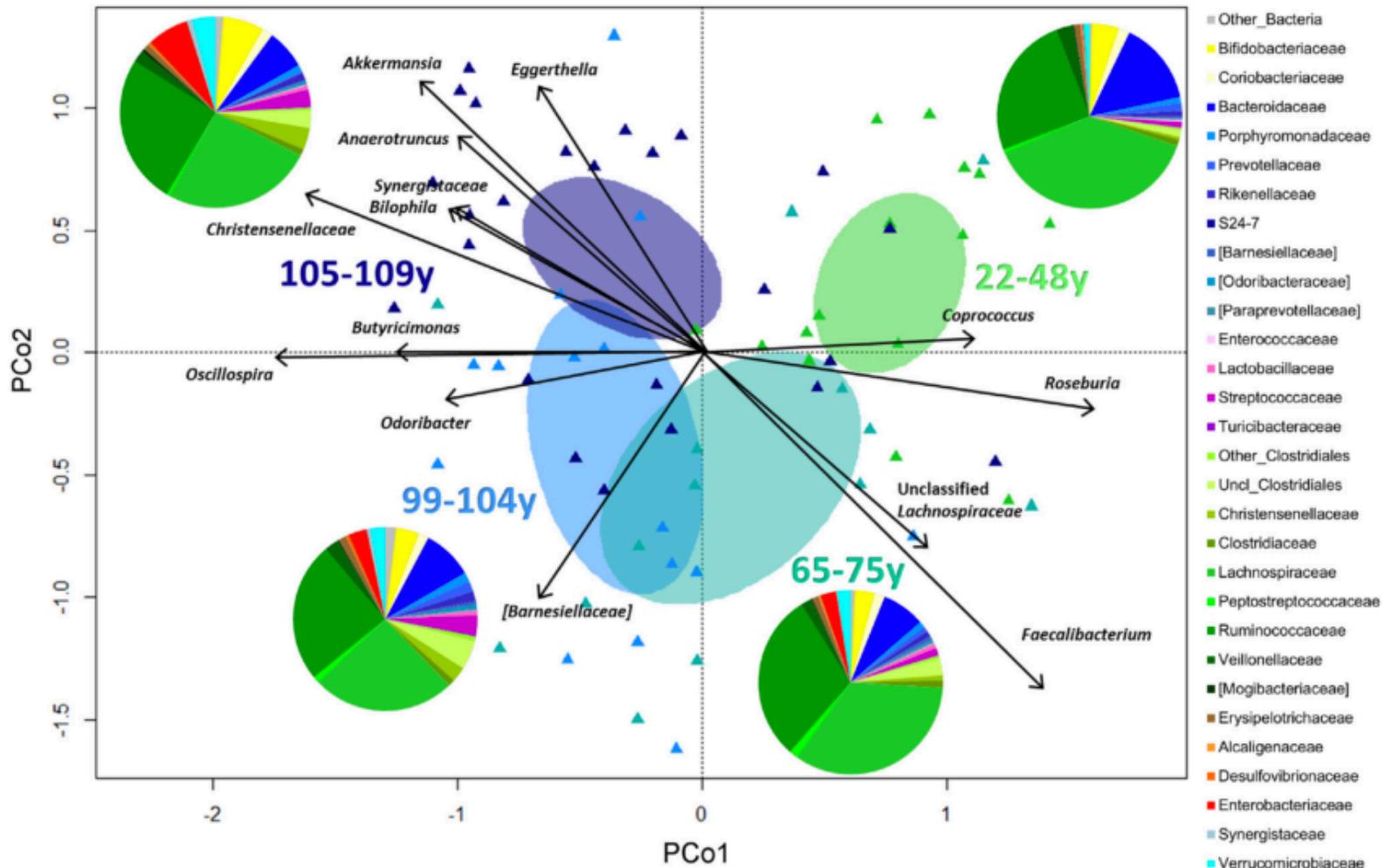
Biagi E, et al: *Through ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians.* PLoS One 2010, 5:e10667.



- Microbial composition and diversity of the gut ecosystem of young adults and seventy-years old people was similar and both differed significantly from that of the centenarians, which showed **lower bacterial diversity**.
- ***Firmicutes* phylum rearrangement in centenarians** with an enrichment in anaerobes, notably **pathobionts**, which associates with increased inflammatory status, with a parallel decrease in *F.prausnitzii* and other bacterial species with anti-inflammatory properties.
- Notably, *E. limosum*, *E. lenta* and *A. Muciniphila* were increased in centenarians.

# Microbiome in Centenarian subjects – Biagi, 2016

Biagi E, et al: Gut Microbiota and Extreme Longevity. Curr Biol  
2016, 26:1480–1485.

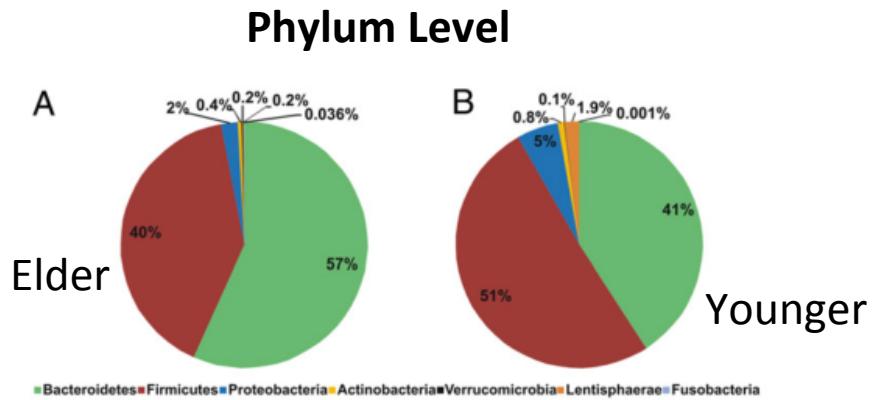


## The microbiome through age

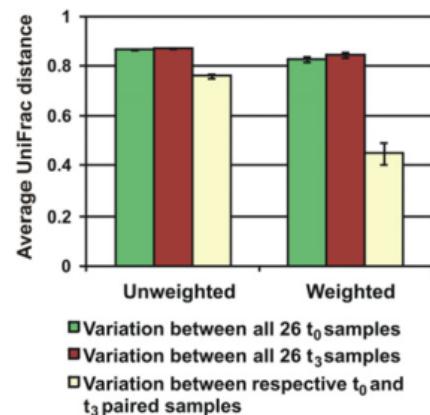
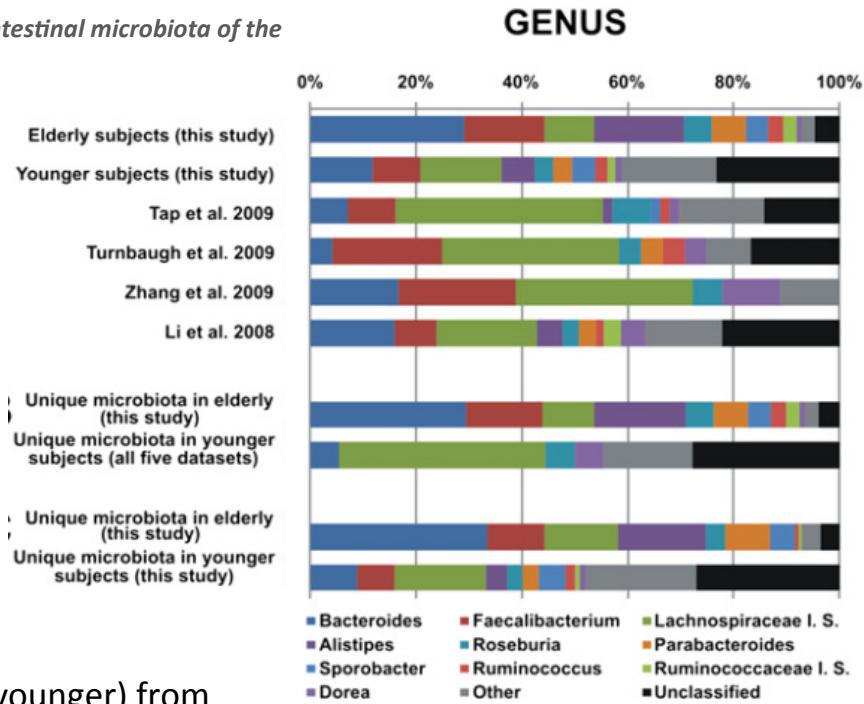


# The Frail Microbiome – Claesson, 2011

Claesson MJ, et al: Composition, variability, and temporal stability of the intestinal microbiota of the elderly. Proc Natl Acad Sci U S A 2011, 108 Suppl:4586–91.



- Cross-Sectional study of 170 individuals (161 >65yo & 9 younger) from ELDERMET consortium. 3-month follow-up of 26 individuals. V3-V4 rRNA gene pyrosequencing.
- Different Bacteroidetes/Firmicutes ratio found in young and elder population highly variable microbiome
- A Bacteroidetes dominated and distinct core microbiome was found for elders whereas younger subjects had one unique Bacteroides species and were Firmicutes phylum dominated in a highly variable context.
- A given microbiota after 3 mo was more like to localize to its original composition than to any other sample analyzed, thus indicating temporal stability



## The Frail Microbiome – Claesson, 2012

Claesson MJ, et al: *Gut microbiota composition correlates with diet and health in the elderly*. Nature 2012, 488:178–184.

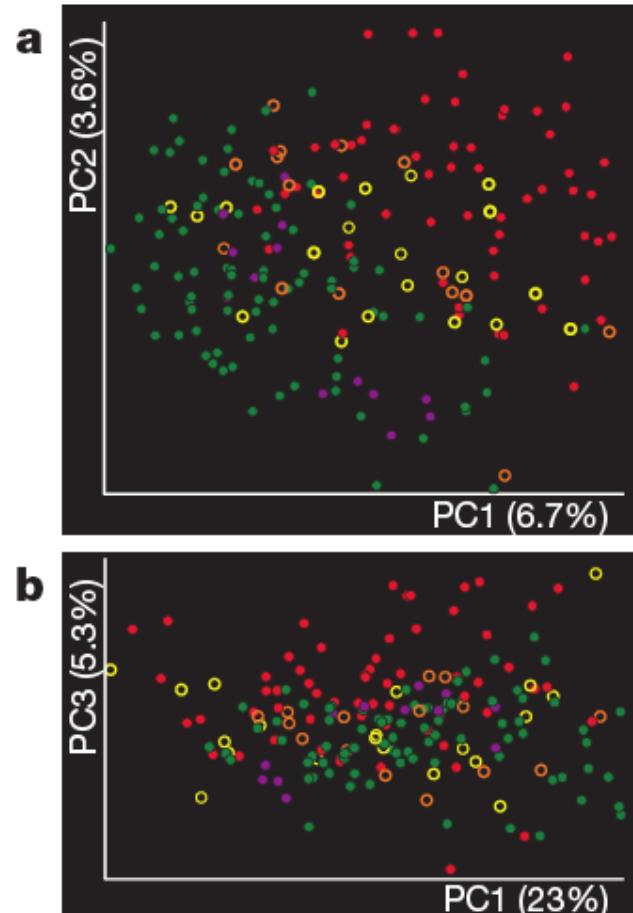
Fecal microbiota characterization of 178 individuals:

- 83 community dwelling subjects (**CD**)
- 20 attending out-patient day hospital (**DH**)
- 15 in short-term rehabilitation (**RH**)
- 60 in long-term residential care, meaning >6 weeks (**LC**)
- 13 young adults (median age 36 y.o.)(**YH**)

Health state characterization

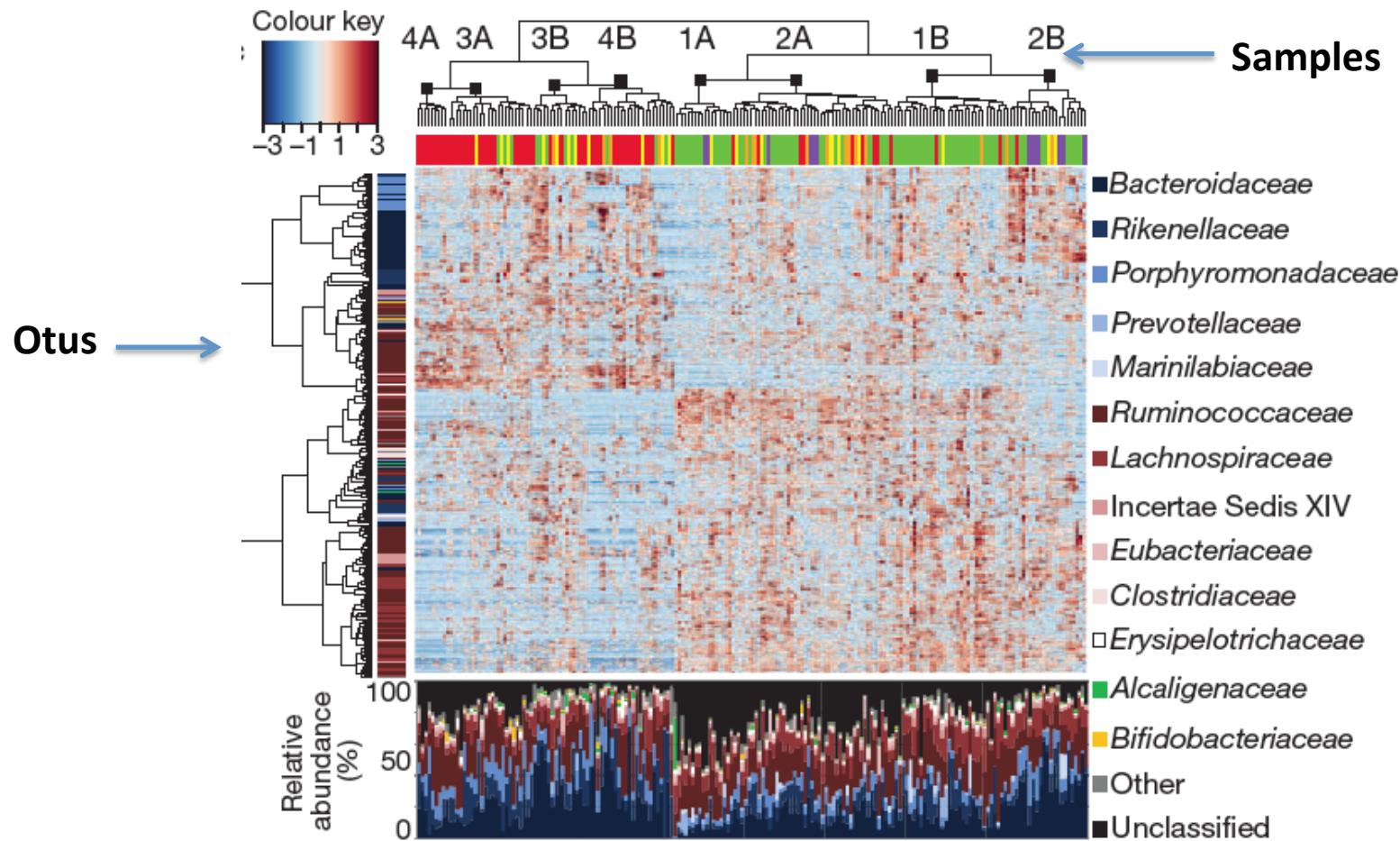
Diet Questionnaires

- The microbiota of older people displays greater inter-individual variation than that of younger adults.
- Clear separation between community-dwelling and long-stay subjects using both weighted and un-weighted analysis
- The faecal microbiota composition from 178 elderly subjects formed groups, correlating with residence location in the community, day-hospital, rehabilitation or in long-term residential care.



Unweighted(a) and Weighted(B) UniFrac PCoA analysis of 47,563 OTUs

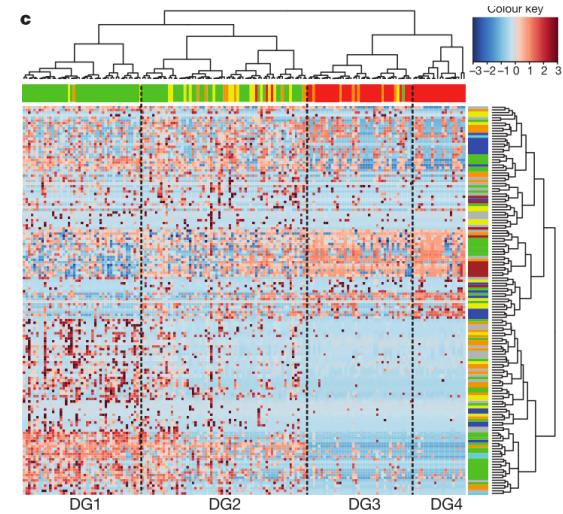
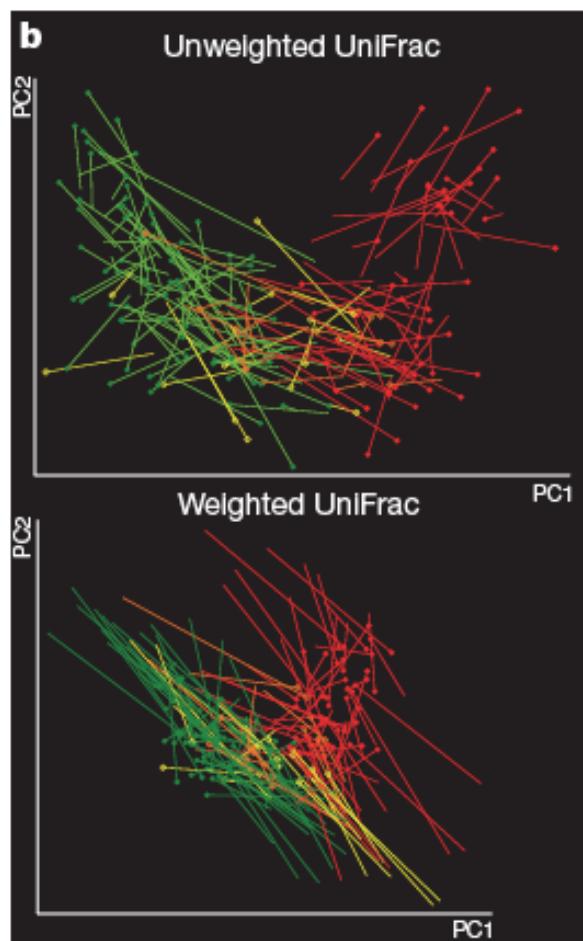
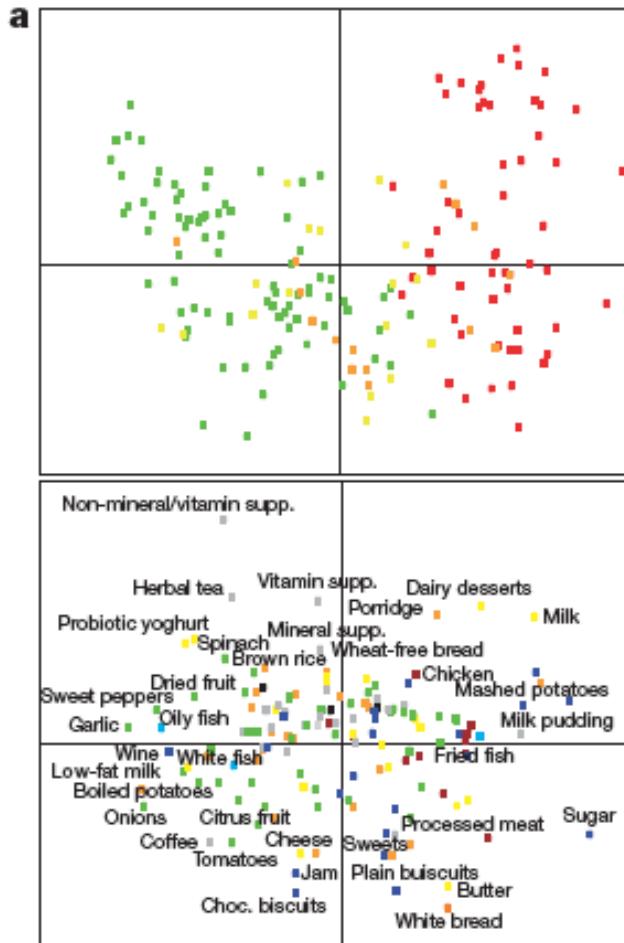
## Microbiota and Residence Location – Claesson, 2012



Hierarchical Ward-linkage clustering based on the Spearman correlation coefficients of the proportion of OTUs, filtered for OTU subject prevalence of at least 20%.

# Microbiome & Diet – Claesson, 2012

Claesson MJ, et al: Gut microbiota composition correlates with diet and health in the elderly. *Nature* 2012, 488:178–184.



**Procrustes analysis: Co-segregation of diet and microbiota along first axis**

CD DH RH LC YH

# Healthy Diet and Microbiota – Claesson, 2012

Claesson MJ, et al: Gut microbiota composition correlates with diet and health in the elderly. *Nature* 2012, 488:178–184.

- Healthy food diversity index (HFD) is based on a distribution measure mainly applied in economic and ecological studies. It considers 3 aspects important for healthy food diversity: number, distribution, and health value of consumed foods

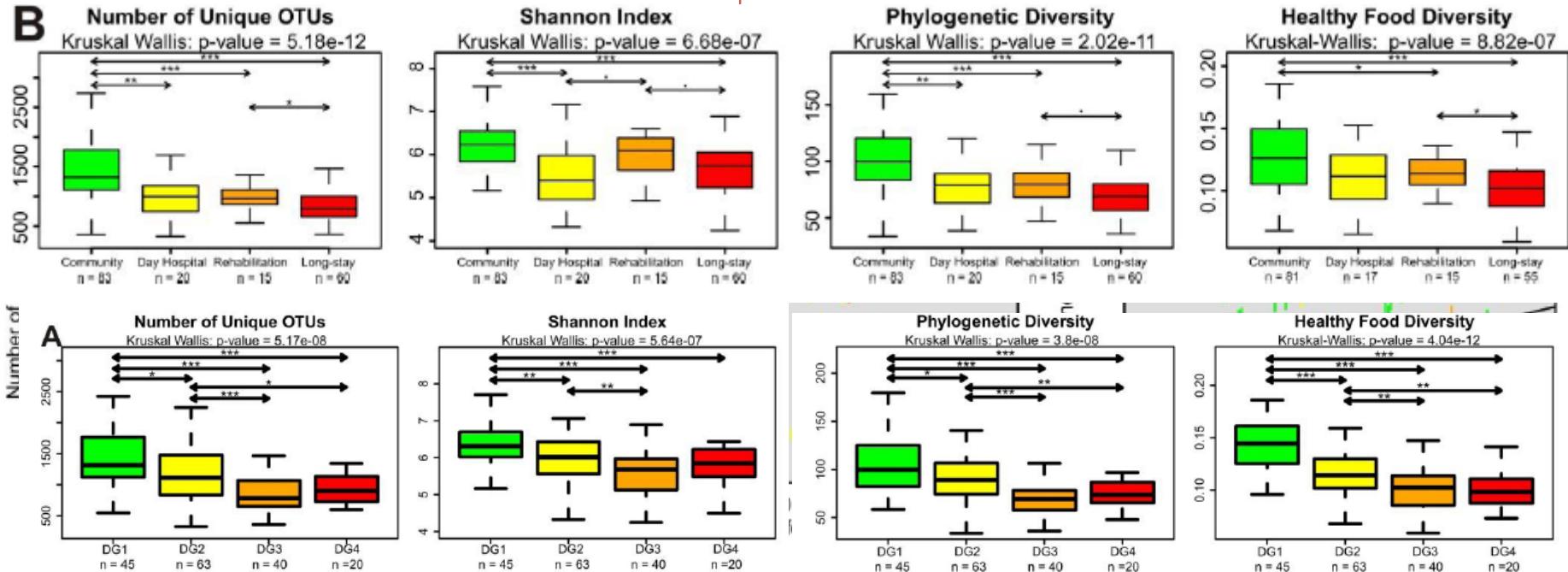
[jn.nutrition.org](http://jn.nutrition.org)  
The Journal of Nutrition  
Nutrient Requirements and Optimal Nutrition 

## A New Index to Measure Healthy Food Diversity Better Reflects a Healthy Diet Than Traditional Measures<sup>1,2</sup>

Larissa S. Drescher,<sup>3</sup> Silke Thiele,<sup>3\*</sup> and Gert B. M. Mensink<sup>4</sup>

<sup>1</sup>Department of Food Economics and Consumption Studies, Christian-Albrechts-University of Kiel, D-24098 Kiel, Germany

and <sup>2</sup>Robert Koch-Institute, D-13353 Berlin, Germany



## Aging Microbiota & Health Correlations – Claesson, 2012

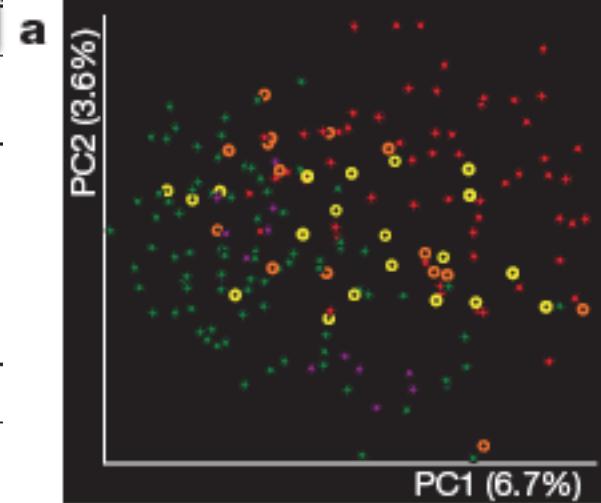
Claesson MJ, et al: *Gut microbiota composition correlates with diet and health in the elderly*. Nature 2012, 488:178–184.

### Regression tests of associations between clinical measurements and microbiota composition (Using PCoA PCs)

| Parameter                | a Unweighted UniFrac PCoA for all four residence locations |              |              |              |              |              |          |
|--------------------------|--|--------------|--------------|--------------|--------------|--------------|----------|
|                          | PC1  |              | PC2          |              | RC range     | RC s.d.      | <i>P</i> |
|                          | RC range   | RC s.d.      | RC range     | RC s.d.      |              |              |          |
| GDT                      | -0.42  | -0.11        | 0.6          | <b>-2.7</b>  | <b>-0.54</b> | <b>0.037</b> |          |
| Diastolic blood pressure | 0.97   | 0.25         | 0.81         | <b>-10.1</b> | <b>-2.02</b> | <b>0.033</b> |          |
| Weight                   | <b>-14.6</b>   | <b>-3.8</b>  | <b>0.033</b> | -7.16        | -1.43        | 0.27         |          |
| CC                       | <b>-3.9</b>  | <b>-1.01</b> | <b>0.022</b> | -2.9         | -0.58        | 0.19         |          |
| IL-6                     | <b>6.71</b>  | <b>1.7</b>   | <b>0.006</b> | <b>6.1</b>   | <b>1.22</b>  | <b>0.007</b> |          |
| IL-8                     | 4.23   | 1.1          | 0.43         | <b>13.6</b>  | <b>2.7</b>   | <b>0.03</b>  |          |
| TNF- $\alpha$            | 1.1  | 0.28         | 0.31         | 0.62         | 0.13         | 0.72         |          |

| Parameter                | b Unweighted UniFrac PCoA for community-only subjects |         |          |             |              |              |          |
|--------------------------|---|---------|----------|-------------|--------------|--------------|----------|
|                          | PC1   |         | PC2      |             | RC range     | RC s.d.      | <i>P</i> |
|                          | RC range  | RC s.d. | RC range | RC s.d.     |              |              |          |
| MNA                      | -1.1  | -0.26   | 0.29     | <b>1.9</b>  | <b>0.5</b>   | <b>0.006</b> |          |
| Diastolic blood pressure | -8.4  | -1.98   | 0.08     | <b>14.3</b> | <b>3.4</b>   | <b>0.035</b> |          |
| GDT                      | -0.13   | -0.03   | 0.8      | <b>-1.5</b> | <b>-0.35</b> | <b>0.02</b>  |          |

| Parameter                | c Unweighted UniFrac PCoA for long-stay-only subjects |             |              |              |              |               |          |
|--------------------------|---|-------------|--------------|--------------|--------------|---------------|----------|
|                          | PC1   |             | PC2          |              | RC range     | RC s.d.       | <i>P</i> |
|                          | RC range  | RC s.d.     | RC range     | RC s.d.      |              |               |          |
| Barthel                  | <b>-6</b>   | <b>-1.5</b> | <b>0.004</b> | <b>-4.8</b>  | <b>-1.3</b>  | <b>0.036</b>  |          |
| FIM                      | <b>-30.8</b>  | <b>-7.8</b> | <b>0.046</b> | <b>-33.3</b> | <b>-4.7</b>  | <b>0.024</b>  |          |
| MMSE                     | -12.15  | -3.08       | 0.14         | <b>-18.4</b> | <b>-4.8</b>  | <b>0.009</b>  |          |
| MNA                      | -3.87   | -0.98       | 0.23         | <b>-11.2</b> | <b>-3</b>    | <b>0.004</b>  |          |
| BMI                      | -1.2  | -0.31       | 0.69         | <b>-5</b>    | <b>-1.3</b>  | <b>0.047</b>  |          |
| CC                       | 0.2   | 0.05        | 0.93         | <b>-6.8</b>  | <b>-1.77</b> | <b>0.0016</b> |          |
| Diastolic blood pressure | <b>19.3</b>   | <b>4.9</b>  | <b>0.015</b> | <b>-12.4</b> | <b>-3.24</b> | <b>0.034</b>  |          |
| Systolic blood pressure  | <b>36.5</b>   | <b>9.3</b>  | <b>0.007</b> | -1.57        | -0.41        | 0.83          |          |
| Weight                   | -3.2  | -0.81       | 0.69         | <b>-12.7</b> | <b>-3.3</b>  | <b>0.024</b>  |          |
| IL-8                     | -2.56   | -0.65       | 0.78         | <b>22.31</b> | <b>5.84</b>  | <b>0.006</b>  |          |
| CRP                      | <b>13.9</b>   | <b>3.53</b> | <b>0.02</b>  | -3.01        | -0.8         | 0.27          |          |

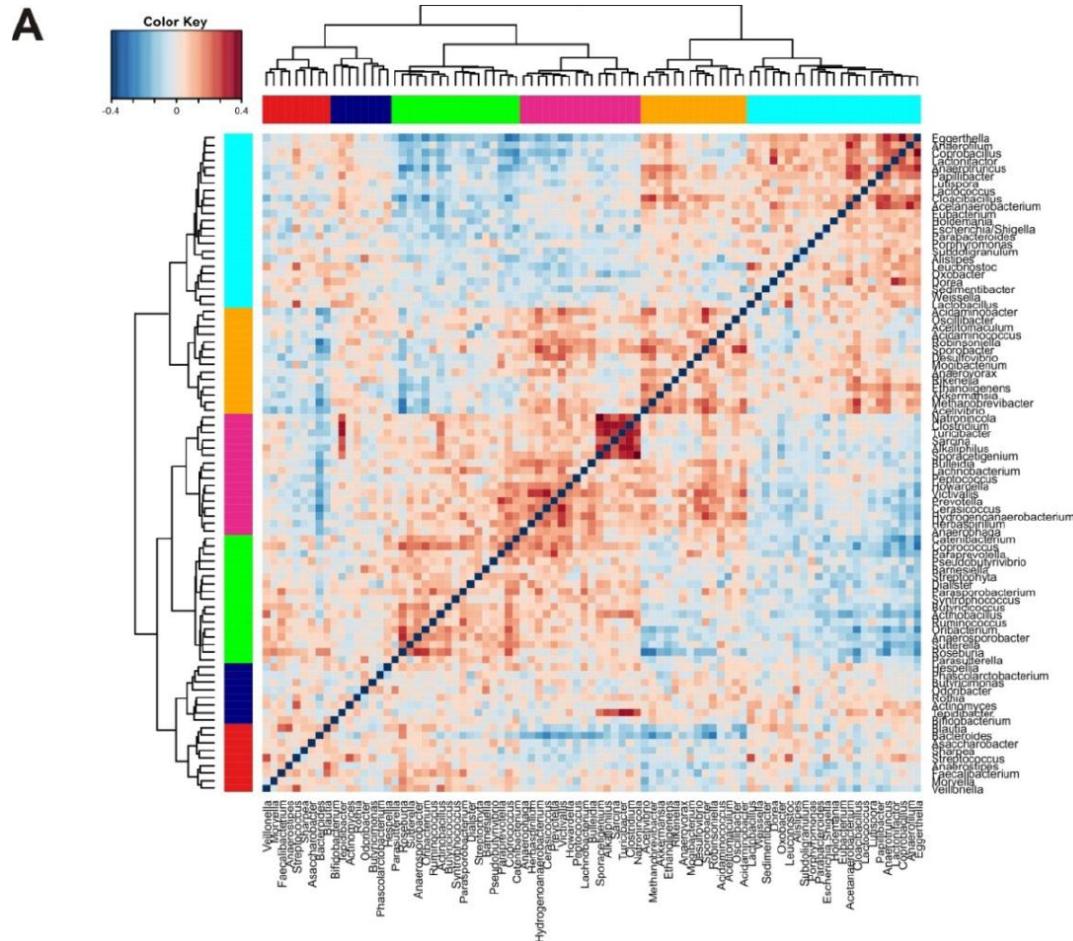


- Charlson Comorbidity index (**CCI**)
- Geriatric depression Test(**GDT**)
- Barthel Index
- Functional Independence Measure (**FIM**)
- Mini-mental state Exam (**MMSE**)
- Mini Nutritional assessment (**MNA**)

## Microbiota structure evolution – Claesson, 2012

Claesson MJ, et al: **Gut microbiota composition correlates with diet and health in the elderly.** Nature 2012, 488:178–184.

**Co-abundance Groups (CAG): Groups of bacteria genera that co-variate along samples**

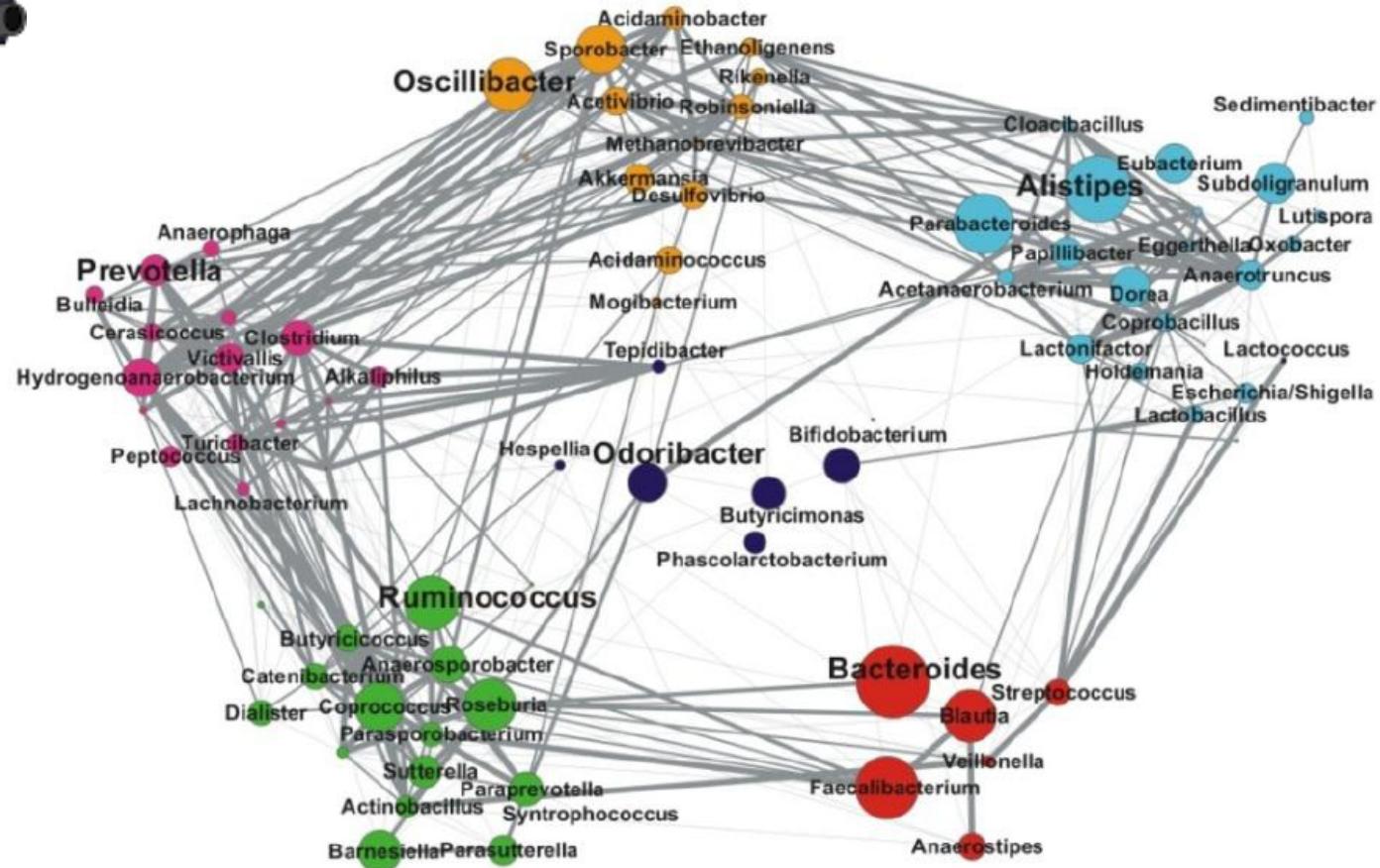


## Microbiota structure evolution – Claesson 2012

Claesson MJ, et al: *Gut microbiota composition correlates with diet and health in the elderly.* Nature 2012, 488:178–184.

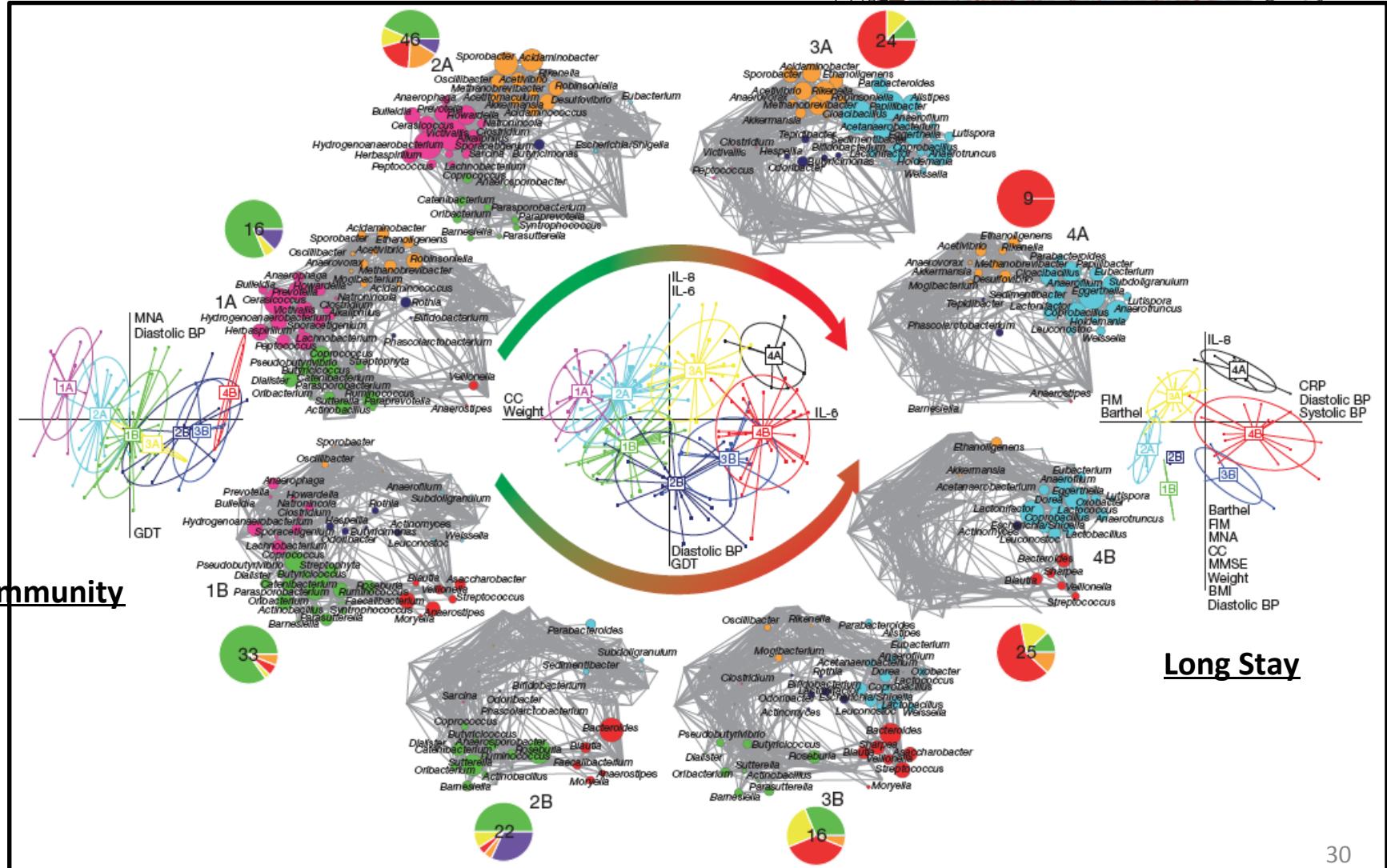
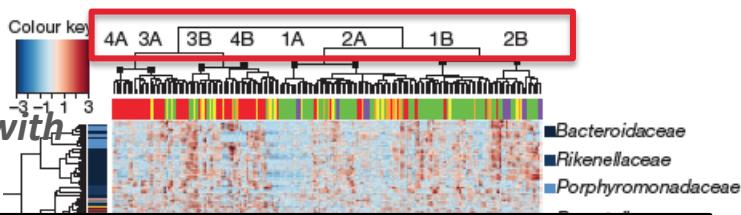


Wiggum Plot



# Microbiota structure evolves through aging and residency – Claesson, 2012

Claesson MJ, et al: Gut microbiota composition correlates with diet and health in the elderly. Nature 2012, 488:178–184.



## Bacterial modules associated to subject characteristics – Jeffery, 2015

Jeffery IB, Lynch DB, O'Toole PW: Composition and temporal stability of the gut microbiota in older persons. ISME J 2015, 10:170–82.

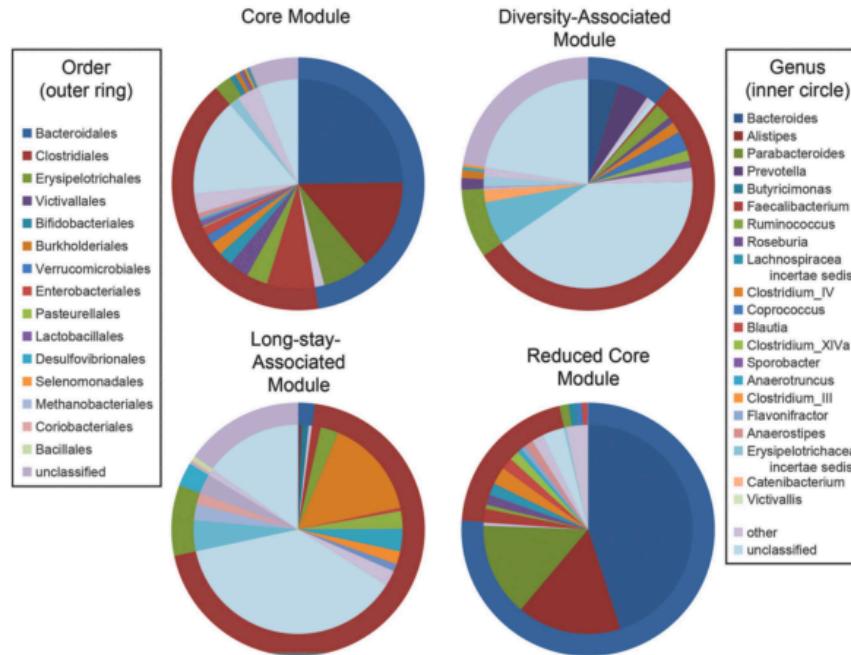
Four microbiota modules identified by iBBiG (M1, M2, M3 and M9) in 732 samples (371 subjects)

**M1:** Core microbiota(Co), bacterial genera present in most microbiome profiles in high abundance

**M2:** Diversity associated (DA), bacterial genera associated with health or healthy diets and high microbiota diversity present in ~1/3 of the profiles

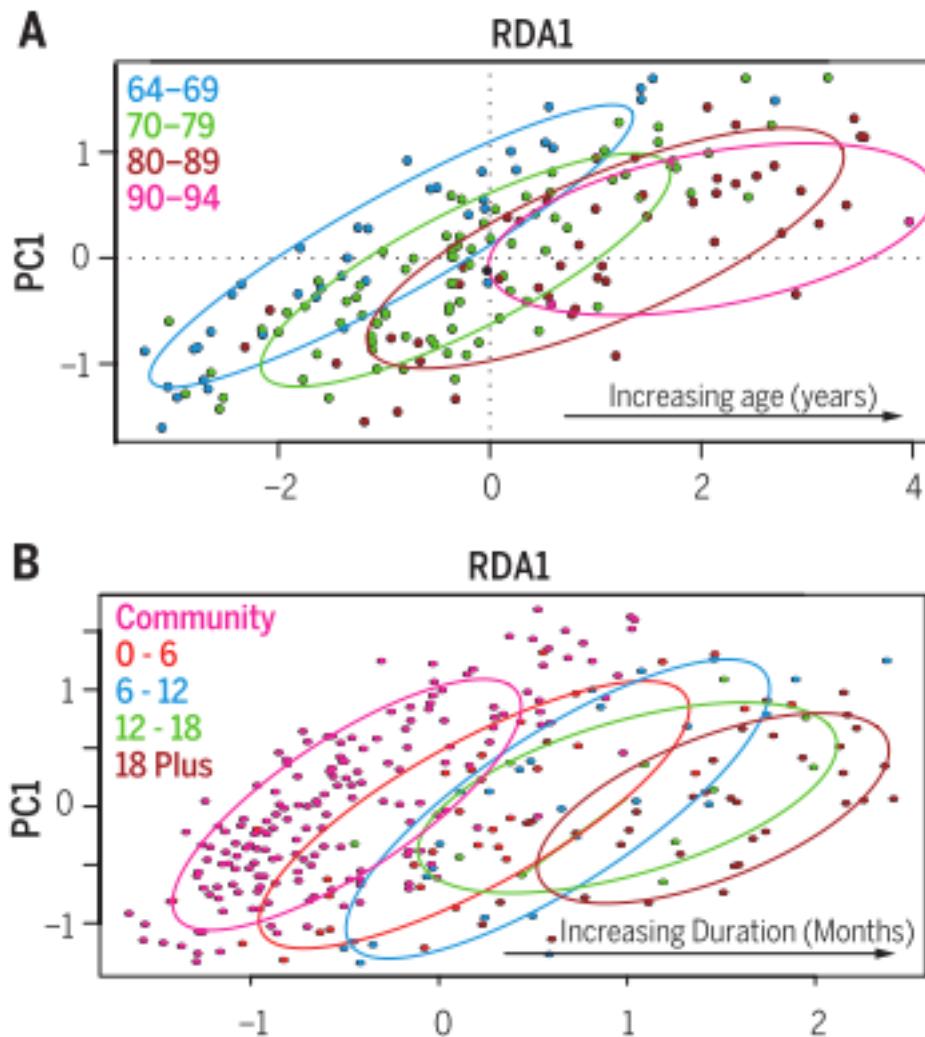
**M3:** Long-stay associated (LA)

**M9:** Reduced Core (RC). A subset of the Co module



## Microbial modules shift through age and residential care – O'Toole, 2015

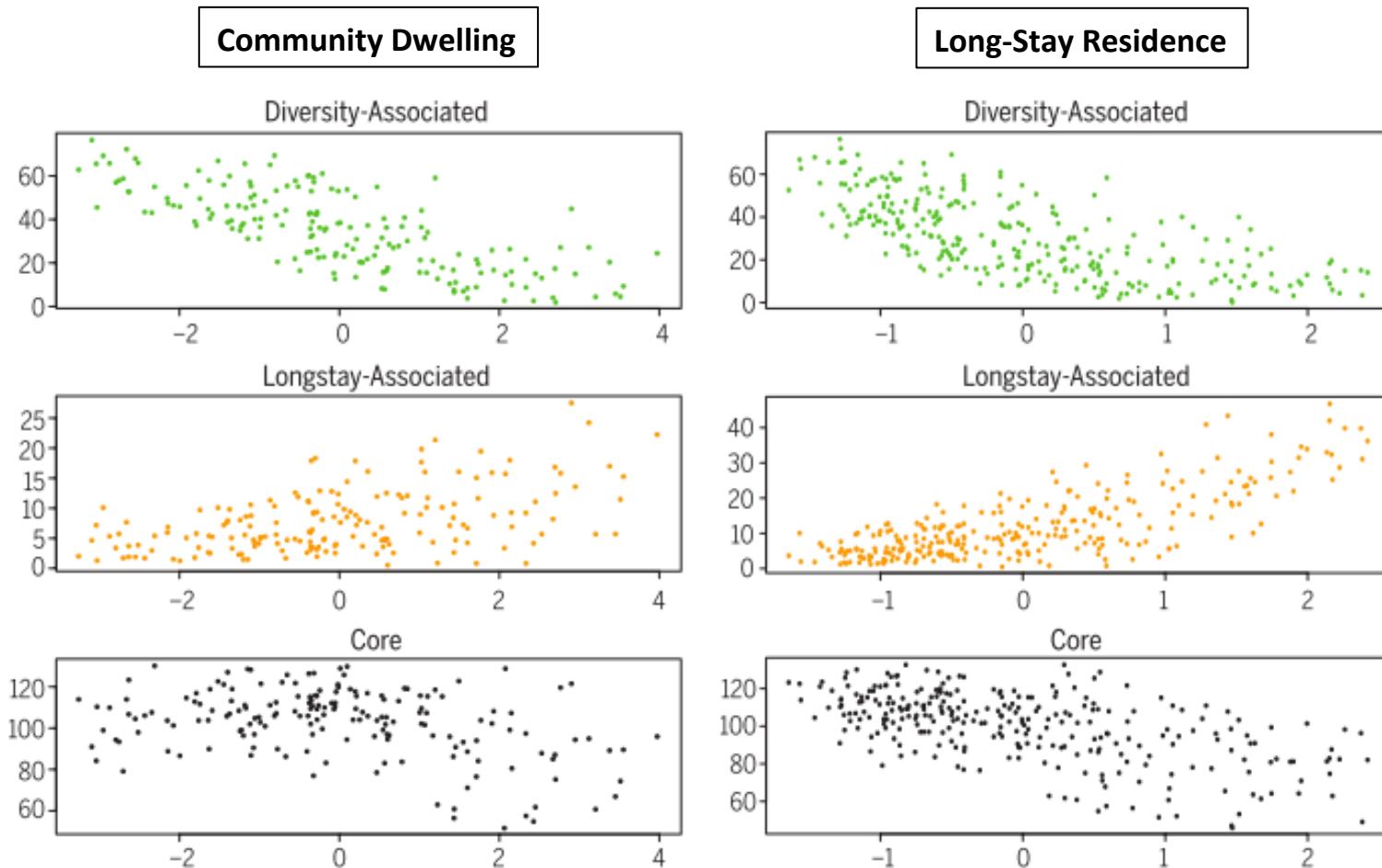
O'Toole PW, Jeffery IB: Gut microbiota and aging. *Science* (80- ) 2015, 350:1214–1216.



## Bacterial modules associated to subject characteristics – O'Toole, 2015

O'Toole PW, Jeffery IB: Gut microbiota and aging. *Science* (80- ) 2015, 350:1214–1216.

- Abundance of bacterial modules with aging and length of stay in residential-care
- As elderly people age and/or spend time in a long-stay residential facility, they gain a long-stay-associated defined population of bacteria that are associated with increased frailty



## Summary

- Measures of increased inflammation and increased frailty related to both dietary and microbiome composition support a diet– microbiota link to these indicators and underlines and important role of environmental factors related to residence setting
- Elder bacterial microbiome gradually differentiates from that of younger population, is stable over time but shows higher inter-individual variability.
- Bacterial diversity is reduced in non-frail aging and is further reduced in frail aging, but comparatively increased in extreme longevity. Frailty is also characterized by reduction of several aging related bacterial components and an increase in *Proteobacteria* and *Enterobacteria*.
- In long living subject, SCFAs producing bacteria such as Akk. *Muciniphila*, *Clostridium XIVa* cluster and *Christensenellaceae* show to be consistently increased compared to “young” population.
- A loss of *F. Prausnitzii* is associated with frailty whereas an enrichment *A. muciniphila* is correlated with longevity, respectively. These species have been shown to be important players in other diseases with an important inflammatory component.
- Importantly, aging/frailty associated microbiome changes are highly population-specific and may depend on a combination environmental factors such as residence, psychosocial factors, diet and presence of co-morbidities.



**The FRAME Study -  
The gut microbiome  
in frail aging**

## Study Design

### Study Population:

- Approx 200 individuals
- 65 yo or older
- With no GI diseases

### Microbiome Characterization

- Sequencing of 16s rRNA V3-V4 region with NGS(Illumina)
- High-Throughput Whole (Meta)Genome sequencing
- Metabolomics: Restricted to SCFA

### Clinical Evaluation:

- Frailty Scales:
  - Fried
  - OARS
  - Barthel
  - Pfeiffer
  - Get up & Go
  - Depression
- Blood & Hormone Test
- Immune and Inflammation markers
  - sCD14 & LBP (Translocation)
  - Zonuline (Gut Perm)
  - IFABP (Enterocyte damage)
  - CRP, D-dimer, IL-6, IP10, G-CSF

### Nutritional Information:

- PREDIMED Questionnaire.
- 14 point scoring.

Screening visit: Consent and material

Baseline Visit: Evaluation and sample collection

### Data Collection:

- On-line data collection
- Secure/Encrypted DB
- OpenClinica data forms designed for this study

## Acknowledgements



### **Microbial genomics**

Roger Paredes  
Chiara Mancuso  
Cristina Rodríguez  
Maria Casadellà  
Marc Noguera-Julian  
Muntsa Rocafort  
Javier Rivera  
Yolanda Guillén  
Mariona Parera



# Thank you!

### **Cell Virology and Immunology**

Jorge Carrillo  
Julià Blanco

Bonaventura Clotet

### **Patients and volunteers**



Pep Coll  
Isabel Bravo  
Cristina Herrero  
Guillem Sirera  
Eugenia Negredo



Javier Santesmases  
Carmen Bracke  
Laura Soldevila  
Cristina Tural



Mari Luz Calle



Ramón Estruch



Domingo Ruiz  
Esther Francia

